# DIFFERENTIAL RESPONSE TO SALINITY IN TWO IRANIAN BARLEY (HORDEUM VULGARE L.) CULTIVARS

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# ABSTRACT

In most southern provinces of Iran, salinity is a growing problem particularly in irrigated agricultural areas with rising water tables, poor water quality and/or deficient soil drainage. To investigate the effects of sodium chloride on two barley cultivars, four levels of salinity: 0, 4, 8 and 12 dS/m, were employed as a factorial experiment arranged in a randomized complete block design with four replications in a controlled environment of the greenhouse during 2008-2009. The results indicated that increasing salinity from 0 to 12 dS/m, decreased the emergence percentage, significantly. The two cultivars (Fajr 30 and Reyhan) responded differently to salinity, Fajr 30 showing a significantly higher emergence rate. This cultivar (Fajr 30) also had greater shoot potassium content. The number of tillers and leaves per plant and also the plant height were decreased upon increasing salinity level. The shoot sodium content was also increased by increasing the salinity level in both cultivars. However, the sodium content of Fajr 30, was lower as compared to Reyhan cultivar, probably due to Na<sup>+</sup> exclusion mechanisms in this cultivar. The results also revealed that the highest grain number and phytomass was obtained from Fajr 30 cultivar at the lowest salinity level. Phytomass and grain yield were significantly decreased at increased salinity. Overall, it appeared that less adverse effect of salinity on Fajr 30 cultivar indicates that this cultivar might be suitable for growing on saline soils.

Key words: salinity, yield components, barley, sodium, potassium.

### **INTRODUCTION**

**C** alt stress is one of the most important abiotic stresses affecting natural productivity causing significant crop loss worldwide. For plants, the sodium ion (Na<sup>+</sup>) is harmful whereas the potassium ion  $(K^{+})$  is an essential ion. The cytosol of plant cells normally contains 100-200 mM of K<sup>+</sup> and 1-10 mM of Na<sup>+</sup> (Taiz and Zeiger, 2002); this  $Na^+/K^+$  ratio is optimal for many metabolic functions in cells. From a physical and chemical point of view,  $Na^+$  and  $K^+$  are similar cations. Therefore, under the typical NaCl-dominated salt environment in nature, accumulation of high Na<sup>+</sup> in the cytosol, and thus higher  $Na^+/K^+$  ratios, disrupts enzymatic functions that are normally activated by K<sup>+</sup> in cells (Bhandal and Malik, 1988; Munns et al., 2006).

Therefore, it is very important for cells to maintain a low concentration of cytosolic  $Na^+$  or to maintain a low  $Na^+/K^+$  ratio in the cytosol when under NaCl stress (Maathuis and Amtmann, 1999).

It has been shown that the two responses occur sequentially, giving rise to a two-phase growth response to salinity (Munns, 1993). For example, comparisons between two genotypes with contrasting rates of Na<sup>+</sup> uptake, and long-term differences in salt tolerance (Schachtman et al., 1991), showed that both genotypes had the same growth reduction for the 4 first weeks in 150 mM NaCl, and it was not until afterwards that a growth difference between the genotypes was clearly observed (Munns et al., 1995). However, within 2 weeks, dead leaves were visible on the more sensitive genotype and the rates of leaf death of old leaves were clearly greater on the sensitive than on the tolerant genotype. Once the number of dead leaves increased above about 20% of the total, plant growth slowed down and many individuals started to die (Munns et al., 1995). Improved salt tolerance of crops can reduce the leaching requirement, and so reduce the costs of an irrigation scheme, both in the need to import fresh water and to dispose of saline water (reviewed by Pitman and Läuchli, 2002). Salt-tolerant crops have a much lower leaching requirement than salt-sensitive ones. In dry-land agriculture, improved salt tolerance can increase yield on saline soils.

In most southern provinces of Iran, where the rainfall is low and the salt remains in the subsoil, increased salt tolerance will allow plants to extract more water. Salt tolerance may have its greatest impact on crops growing on soils with natural salinity as, when all the other agronomic constraints have been overcome (e.g. disease attacks and nutrient deficiency); subsoil salinity remains a major limitation to agriculture in all semi-arid regions, such as most southern provinces of Iran. Even where clearing of land in higher rainfall zones caused water-tables to rise and salt to move, improved salt tolerance of crops will have a place. The introduction of deeprooted perennial species can be useful in order to lower the water-table, however, salt tolerance will be required not only for the 'dewatering' species, but also for the annual crops that follow, as salt will be left in the soil when the water-table is lowered (Francois et al., 1994).

Barley is a relatively tolerant crop to soil salinity, and genetic variations exist among genotypes of cultivated barley. One of the two new cultivars of barley, used in the present study, Fajr 30, is an improved hybrid recommended for Salinity areas in most southern provinces of Iran (Pakniyat et al., 2003). Fair 30 barley cultivar has been released based on its high grain yield and vield stability as well as its desirable agronomic characteristics which have contributed to its wide adaptation in temperate areas of Iran, However, the salt tolerance mechanisms of this variety has not been studied in detail. The objective of the present study was to quantify plant growth, yield and yield components of the two barley cultivars in relation to various concentrations of NaCl. In addition, NaCl effects on the chemical composition of the plant organs were measured. The experiment was carried out in the Fars province, one of the main barleygrowing areas in southern Iran, with more than 430,000 ha barley grown as nearly continuous cropping.

# **MATERIAL AND METHODS**

# Site, treatment application and data collection

This experiment was conducted to evaluate the effects of sodium chloride on two barley cultivars (Fajr 30, a relatively salt tolerant genotype and Reyhan, a salt sensitive cultivar) and four levels of salinity: 0, 4, 8 and 12 dS/m. Fajr 30 barley cultivar is moderately susceptible to powdery mildew, resistant to lodging, and tolerant to low temperatures and drought. Irrigated barley is grown on about 320,000 hectares in the temperate agroclimatic zone of Iran. The average grain yield of barley in these areas is 3300 kg/ha. New improved high yielding cultivars and appropriate agronomic practices is a logical strategy to increase the grain yield and production of irrigated barley in the temperate agro-climatic zone of Iran. Fajr 30 barley cultivar was released in 2009 by the Seed and Plant Improvement Institute, based on its high grain yield and yield stability, as well as its desirable agronomic characteristics which have contributed to its wide adaptation in temperate areas of Iran.

The desired salinity levels were obtained by mixing the required amount of NaCl and  $CaCl_2$  (5:1) in soil before filling the pots (0, 2.16, 4.32, 8.64 g/kg soil). The barley crop was sown on 10 November 2008 and harvested on 30 April 2009. The experiment was carried out in a greenhouse at the college of agriculture, Shiraz university, Shiraz Iran (52°46'E, 29°50'N, altitude 1810 m asl), 12 km north of Shiraz, on a Fine mixed, mesic Typic Calcixerpets soil with air temperature in the range of about 25 to 30°C and light intensity in the range of about 600-1000  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, and was conducted using as a factorial experiment arranged in a randomized complete block design with four replications. Soil properties are shown in table 1. Pregerminated seeds were sown in 5 L perforated plastic pots filled with fertilized (50, 25 and 25 N, P and K mg kg<sup>-1</sup>, respectively) and were kept in concrete tanks filled with tap water according to Maas (1985). The level of water was maintained at 3 cm below the soil surface for 2 days. Ten seeds of each cultivar were sown in each pot, thinned to five seedlings at two-leaf stage. The pots were kept flooded thereafter for the rest of the experiment. The emergence percentage and number of leaves per plant were recorded throughout the experiment. Plants were harvested and threshed manually. The data regarding grain number, straw yield and grain weight, number of spikes per plant, number of tillers per plant and shoot length were recorded (Wilhelm et al., 1989).

Year	OC (%)	pН	Sand (%)	Silt (%)	Clay (%)	Soil texture	EC (dSm <sup>-1</sup> )	P (mg kg <sup>-1</sup> )	$\frac{K}{(mg kg^{-1})}$	Total N (%)
2008-2009	0.73	7.1	7.1	66.6	26.3	Silty loam	0.03	15.5	376	0.07

#### Sodium and potassium measurements

Dried samples at the harvesting date were ground to a fine powder and about 0.1 g was transferred to a test tube containing 10 mL of 0.1 N acetic acid, and heated in a water bath at 80 °C for 2 h. The extracted tissue was cooled at room temperature and left overnight, and then filtered using Whiteman filter paper number 40. Sodium and potassium concentrations were then determined using an atomic absorption spectrometer (Munns and James, 2003).

# **Proline measurements**

Fresh flag leaf tissue (0.5 g) was ground in liquid nitrogen and then extracted in 20 ml of hot water for 30 min with moderate shaking. The homogenate was centrifuged at 5000 g for 10 min. The proline concentration was quantified using the ninhydrin acid reagent method as described by Bates et al. (1973) using L-proline as a standard.

# Statistical analysis

Statistical analysis was performed for each parameter studied based on a randomized complete block design model with four replications using SAS software (SAS Inst., 1985). Means were separated by Duncan's Multiple Range Tests at  $p \le 0.05$ .

#### RESULTS

The results indicated that increasing salinity from 0 to 12 dS/m, significantly decreased emergence percentage. The two cultivars (Fajr 30 & Reyhan) responded

differently to salinity, so that Fajr30 showed significantly higher emergence rate. The results revealed that the highest grain number and phytomass was obtained from Fajr 30 cultivar at the lowest salinity level (Table 3). Phytomass and grain yield were significantly decreased at increased salinity.

Our results showed that Fair 30 cultivar had higher shoot potassium concentration (Table 4). There is a strong correlation between salt exclusion and salt tolerance in many species (reviewed by Läuchli, 1984; Munns and James, 2003). Figure 1 shows the negative relationship between leaf Na<sup>+</sup> concentration and salt tolerance of Fair 30 cultivar, measured as biomass under salinity stress expressed in % of control. In general; the Fajr 30 cultivar compared to Reyhan, had lower Na<sup>+</sup> concentrations at all salinity levels (Table 4). The results showed that there was a significant difference among different salinity levels for proline content of the two cultivars, and Fajr 30 cultivar had higher proline content (Table 4).

### DISCUSSION

# Effect of different sodium chloride levels on growth and morphological characteristics

Experimental treatments had significant effects on morphological traits of both cultivars. The number of tillers and leaves per plant and also the plant height were decreased upon increasing salinity level (Table 2), which is in agreement with the finding of Abdullah et al. (1978). Fajr 30 was superior to Reyhan as far as the salinity tolerance characteristics (as shown in Table 2) were concerned. Kingsbury et al. (1984) showed that the major difference between two lines of barley in salinity tolerance was their response to specific ion effects, at the level of the organ, tissue, cell, and sub-cellular entities. Superior compartmentalization of toxic ions by the more salt-tolerant line, presumably in the vacuole, might have enabled it to maintain its cytoplasmic metabolic apparatus in a stable and more nearly normal state than the sensitive line. Therefore, a measure of true cytoplasmic toleration of salt may also be needed to be considered as a factor. The first phase of the growth response results from the effect of salt outside the plant i.e. the salt in the soil solution (the osmotic stresses), which reduces leaf growth, as shown in table2.

Indeed, salts themselves do not build up in the growing tissues at concentrations that inhibit growth, as the rapidly elongating cells can accommodate the salt that arrives in the xylem within their expanding vacuoles. So, the salt taken up by the plant does not directly inhibit the growth of new leaves (Munns, 1993).

Treatments	Emergence percent	Leaves per plant	Tillers per plant	Plant height (cm)	Spikes per plant
Cultivars	•	•	1		· · ·
(V <sub>1</sub> ) Reyhan	52.58 a	5.11 a	1.13 a	30.26 a	1.16 a
(V <sub>2</sub> ) Fajr 30	61.41 a	7.06 a	1.69 a	32.06 a	1.31 a
Salinity (dS/m)					
$(S_0) = 0$	93.00 a	13.23 a	3.00 a	52.17 a	2.40 a
$(S_1)$ 4	93.57 a	10.10 b	2.50 a	42.67 b	1.53 b
(S <sub>2</sub> ) 8	54.00 b	3.23 c	1.16 b	27.50 c	0.73 c
(S <sub>3</sub> ) 12	3.23 c	_*	-	-	-
$V_1S_0$	91.67 a	13.13 a	2.06 ab	47.33 ab	2.13 ab
$V_1S_1$	92.00 a	8.23 b	2.10 bc	47.33 ab	1.26 bc
$V_1S_2$	45.67 c	2.10 cd	0.46 d	24.33 d	0.16 de
$V_1S_3$	0.00 e	0.00 d	0.00 d	0.00 d	0.00 d
$V_2S_0$	94.33 a	13.33 a	3.13 a	57.00 a	2.16 a
$V_2S_1$	92.33 a	11.57 a	3.00 a	40.00 bc	2.20 ab
$V_2S_2$	62.33 b	4.32 c	1.46 c	30.67 cd	1.10 cd
$V_2S_3$	6.16 d	-	-	-	-

Table 2. Comparison of main and interaction effects of morphological traits

Means at each column for each character, followed by similar letters are not significantly different using Duncan's multiple range tests ( $p \le 0.05$ ). \*No plant growth due to salinity.

The second phase of the growth response results from the toxic effect of salt inside the plant. The salt taken up by the plant concentrates in the old leaves; continued transport of salt into transpiring leaves over a long period of time eventually results in very high Na<sup>+</sup> and  $Cl^-$  concentrations, and the leaves die as it was observed in our experiment (Tables 2 and 4). The cause of the injury is probably due to the salt load exceeding the ability of the cells to compartmentalize salts in the vacuole. Salts then would rapidly build up in the cytoplasm and inhibit enzyme activity (Munns, 1993). Alternatively, they might build up in the cell walls and dehydrate the cell (Flowers et al., 1991). However, Mühling and Läuchli (2002) found no evidence for this in maize cultivars that differed in salt tolerance.

# Relationship between salinity and yield components

Phytomass and grain yield were also significantly decreased by salinity. Yield reduction was attributed primarily to reduced spike weight and individual seed weight, rather than spike number (Table 3). This finding confirms the results of Francois et al. (1989). The straw yield was more sensitive to salinity than was the grain yield (Table 3). Our results also suggest that estimates of grain yield might bring another complexity to the salinity response, not just because the crops must be grown in controlled environments for long periods of time, but also due to complexity of the conversion of shoot biomass to grain. A low level of salinity may not reduce grain weight even though the leaf area and phytomass is reduced (Table 3). The grain yield may not decrease until a given ('threshold') salinity is reached (Maas and Hoffman, 1977).

Treatments	No of grains per plants	Grains weight per plant (g)	Grain yield per plant (g)	Phytomass (g)	Leaf area at anthesis (cm <sup>2</sup> )	Straw weight (g)	Spike weight (g)
Cultivars							
(V <sub>1</sub> ) Reyhan	10.25 a	0.19 a	1.55 b	3.11 b	4600 a	1.28 b	2.70 b
(V <sub>2</sub> ) Fajr 30	12.16 a	0.18 a	2.25 a	4.01 a	3700 b	1.45 a	3.50 a
Salinity (dS/m)							
$(S_0)  0$	19.17 a	0.33 a	8.04 a	11.57 a	4900 a	3.07 a	10.20 a
$(S_1)$ 4	15.00 ab	0.35 a	3.55 b	6.11 b	3950 b	2.11 b	5.65 b
(S <sub>2</sub> ) 8	10.67 b	0.03 b	0.33 c	1.26 c	2800 c	0.86 c	0.88 c
(S <sub>3</sub> ) 12	_*	-	-	-	-	-	-
$V_1S_0$	14.00 ab	0.44 a	6.26 a	10.25 a	4350 a	2.45 b	10.15 a
$V_1S_1$	14.33 ab	0.21 bc	2.66 b	5.11 b	2500 ab	2.15 b	4.51 b
$V_1S_2$	10.67 bc	0.01 c	0.12 c	0.59 c	1900 d	0.43 d	0.34 c
$V_1S_3$	-	-	-	-	-	-	-
$V_2S_0$	24.33 a	0.13 ab	7.03 a	12.49 a	4750 a	3.43 a	11.29 a
$V_2S_1$	15.67 ab	0.32 abc	4.50 b	8.18 b	4210 b	2.18 b	6.48 b
$V_2S_2$	10.67 bc	0.05 bc	0.35 c	3.11 c	2700 с	1.19 c	1.21 c

Table 3. Comparison of main and interaction effects on yield and yield components of two barley cultivars

Means at each column for each character, followed by similar letters are not significantly different using Duncan's multiple range tests ( $p \le 0.05$ ). \*No plant growth due to salinity.

# Effect of different sodium chloride levels on chemical composition

The shoot sodium concentration was also increased by increasing the salinity level in both cultivars; however, the sodium content of Fajr 30 cultivar, compared to Reyhan, was lower, probably due to Na<sup>+</sup> exclusion mechanisms in this cultivar (Table 4). The increase in Na<sup>+</sup> and Cl<sup>-</sup> and decrease in K<sup>+</sup> contents of barley grains suggest that the effect of salinity on the physiological phenomenon studied is due to changes in the ionic content of the plants (Abdullah et al., 1978). Salt tolerance in barley can be improved based on mechanisms for salt tolerance, using physiological traits to select germplasm. In barley, salt tolerance is associated with low rates of transport of Na<sup>+</sup> to shoots, with high selectivity for  $K^+$  over Na<sup>+</sup> (Gorham et al., 1987, 1990). Correlations between grain yield and Na<sup>+</sup> exclusion from

leaves, along with the associated enhanced  $K^+/Na^+$  discrimination, was shown in barley (Chhipa and Lal, 1995; Ashraf and O'Leary, 1996; Ashraf and Khanum, 1997), although the relationship may not hold across all genotypes (Ashraf and McNeilly, 1988; El-Hendawy et al., 2005). This suggests that Na<sup>+</sup> exclusion is not the only mechanism of salt tolerance (Colmer et al., 2006).

There is a strong correlation between salt exclusion and salt tolerance in many species (reviewed by Läuchli, 1984; Munns and James, 2003). Figure 1 shows the negative relationship between leaf Na<sup>+</sup> concentration and salt tolerance of Fajr 30 cultivar. In general, the Fajr 30 cultivar had lower Na<sup>+</sup> concentrations as compared to Reyhan and produced more dry matter (Table 4). This low-Na<sup>+</sup> genotype had fewer injured leaves, and a higher proportion of living to dead leaves, as observed during the experiment. The effect on growth was probably due to a better carbon balance in the genotype with less  $Na^+$ . A similar relationship between shoot dry matter and leaf  $Na^+$  was found in a population from a cross between high- and low-Na<sup>+</sup> genotypes (Munns and James, 2003). The proline content was also increased in both cultivars by increasing the salinity level (Table 4).



*Figure 1*. Relationship between leaf Na<sup>+</sup> concentration and salinity tolerance (measured as biomass in % of control) in Fajr 30 cultivar ( $R^2 = 0.752$ ). Na<sup>+</sup> concentrations were measured on leaf 3 after 10 d and shoot biomass after 24 d in 150 mM NaCl. Values are expressed as a percentage of shoot biom in control conditions. All values are means (n = 5).

Treatments	K <sup>+</sup> (mmol per Kg)	Na <sup>+</sup> (mmol per Kg)	Proline (µ g/g)
Cultivars			
(V <sub>1</sub> ) Reyhan	210.70 b	15.10 a	0.24 b
(V <sub>2</sub> ) Fajr 30	410.50 a	13.70 b	0.32 a
Salinity (dS/m)			
$(S_0)  0$	319.40 c	94.10 d	0.25 d
(S <sub>1</sub> ) 4	410.70 b	87.30 b	0.27 b
(S <sub>2</sub> ) 8	586.50 a	160.50 a	0.41 a
(S <sub>3</sub> ) 12	-	-	-
$V_1S_0$	287.20 d	141.14 d	0.25 d
$V_1S_1$	209.00 d	168.80 b	0.26 b
$V_1S_2$	394.90 c	318.40 a	0.33 a
$V_1S_3$	-	-	-
$V_2S_0$	351.70 c	46.80 de	0.29 d
$V_2S_1$	612.30 b	5.80 e	0.30 ab
$V_2S_2$	778.10 a	2.50 e	0.37 a

*Table 4*. Comparison of main and interaction effects on chemical composition of two barley cultivars

Means at each column for each character, followed by similar letters are not significantly different using Duncan's multiple range tests ( $p \le 0.05$ ).

\*No plant growth due to salinity.

Moradi and Ismail (2007) reported that it has been repeatedly inferred, but not yet proven, that there might be a relationship between salt tolerance and the accumulation of proline and other metabolites for osmotic adjustment. However, Colmer et al. (1995) suggested that the increase in proline concentration may not be associated with salinity tolerance. Indeed, elevated proline levels may also confer additional regulatory or osmoprotective functions under salt stress, such as the control of the activity of plasma membrane transporters involved in cell osmotic adjustment in barley roots (Cuin and Shabala, 2005).

## CONCLUSIONS

Our results indicated that the two cultivars (Fajr 30 and Reyhan) responded differently to salinity, so that Fajr 30 showed significantly higher emergence rate. This cultivar (Fajr 30) also had higher shoot potassium content. The number of tillers and leaves per plant and also the plant height were decreased in both cultivars by increasing salinity level.

The shoot sodium content was also increased by increasing the salinity level in both cultivars; however, the sodium content of Fajr 30 cultivar, compared to Reyhan, was lower, probably due to  $Na^+$  exclusion mechanisms in this cultivar. The results also revealed that the highest grain number and phytomass was obtained from Fajr 30 cultivar at the lowest salinity level. Phytomass and grain yield were also decreased upon salinity significantly. Fajr 30, which is a tolerant cultivar, originates from Ardekan (yazd) which is a dry, saline area in the central part of Iran. Therefore, it may conclude that, not surprisingly, harsh environments due to salinity may result in natural selection of tolerant genotypes. Overall, it appeared that less adverse effects of salinity on Fajr30 cultivar may make it more suitable for growth in saline soils. This subject is worth further exploring.

#### REFERENCES

- Abdullach, Z., Ahmad, R., Ahmad, J., 1978. *Salinity induced changes in the reproductive physiology of wheat plants.* Plant and Cell Physiol., 19: 99-106.
- Ashraf, M., Khanum, A., 1997. Relationship between ion accumulation and growth in two spring wheat lines differing in salt tolerance at different growth stages. Journal of Agronomy and Crop Science, 178: 39-51.
- Ashraf, M., McNeilly, T., 1988. Variability in salt tolerance of nine spring wheat cultivars. Journal of Agronomy and Crop Science, 160: 14-21.
- Ashraf, M., O'Leary, J.W., 1996. Responses of some newly developed salt-tolerant genotypes of spring wheat to salt stress. 1. Yield components and ion distribution. Journal of Agronomy and Crop Science, 176: 91-101.
- Bates, L.S., Waldren, R.P., Teare, I.D., 1973. Rapid determination of free proline for water-stress studies. Plant Soil., 39: 205-207.
- Bhandal, I.S, and Malik, C.P., 1988. *Potassium* estimation, uptake, and its role in the physiology and metabolism of flowering plants. International Review of Cytology, 110: 205-254.
- Chhipa, B.R., Lal, P., 1995. *Na/K ratios as the basis of salt tolerance in wheat*. Australian Jouranl of Agricultural Research, 46: 533-539.
- Colmer, T.D., Epstein, E., Dvorak, J., 1995. Differential solute regulation in leaf blades of various ages in salt-sensitive wheat and salt tolerant wheat x Lophopyrum elongatum (Host) A. Löve amphiploid. Plant Physiology, 108: 1715-1724.
- Colmer, T.D., Flowers, T.J., Munns, R., 2006. Use of wild relatives to improve salt tolerance in wheat. Journal of Experimental Botany, 57: 1059-1078.
- Cuin, T.A., Shabala, S., 2005. Exogenously supplied compatible solutes rapidly ameliorate NaClinduced potassium efflux from barley roots. Plant and Cell Physiology, 46: 1924-1933.

- El-Hendawy, S.E., Hu, Y., Schmidhalter, U., 2005. Growth, ion content, gas exchange, and water relations of wheat genotypes differing in salt tolerances. Australian Journal of Agricultural Research, 56: 123-134.
- Flowers, T.J., Hajibagheri, M.A., Yeo, A.R., 1991. Ion accumulation in the cells walls of rice plants growing under saline conditions: evidence for the Oertli hypothesis. Plant, Cell and Environment, 14: 319-325.
- Francois, L.E., Donovan, T.J., Lorenz, K. and Maas, E.V., 1989. Salinity Effects on Rye Grain Yield, Quality Vegetative Growth, and Emergence. Agron. J., 81:707-712.
- Francois, L.E., Grieve, C.M., Maas, E.V., Donovan, T.J., Lesch, S.M., 1994. *Time of salt stress affects* growth and yield components of irrigated wheat. Agron. J., 86: 100-107.
- Gorham, J. Hardy, C., Wyn Jones, R.G., Joppa, L.R., Law, C.N., 1987. Chromosomal location of a K/Na discrimination character in the D genome of wheat. Theorertical and Applied Genetics, 74: 584-588.
- Gorham, J., Wyn, Jones, R.G., Bristol, A., 1990. Partial characterization of the trait for enhanced  $K^+$  -  $Na^+$  discrimination in the D genome of wheat. Planta, 180: 590-597.
- Kingsbury, Ralph, W., Epstein, E. and Pearcy, W., 1984. *Physiological Responses to Salinity in Selected Lines of Wheat*. Plant Physiology, 74: 417-423.
- Läuchli, A., 1984. Salt exclusion: an adaptation of legumes for crops and pastures under saline conditions. In: Staples R.C., (ed.), Salinity tolerance in plants: strategies for crop improvement, New York, NY: Wiley: 171-187.
- Maas, E.V, Hoffman, G.J., 1977. Crop salt tolerance current assessment. Journal of the Irrigation and Drainage Division of the American Society of Civil Engineering, 103: 115-134.
- Maas, E.V., 1985. Crop tolerance to saline sprinkling water. Plant Soil, 89: 273-284.
- Maathuis, F.J.M., and Amtmann, A., 1999.  $K^+$  nutrition and  $Na^+$  toxicity the basis of cellular  $K^+/Na^+$ ratios. Annals of Botany, 84: 123-133.
- Moradi, F., and Ismail, M., 2007. Responses of Photosynthesis, Chlorophyll Fluorescence and ROS-Scavenging Systems to Salt Stress during Seedling and Reproductive Stages in Rice. Annals of Botany .99(6): 1161-1173.
- Mühling, K.H, and Läuchli, A., 2002. Effect of salt stress on growth and cation compartmentation in leaves of two plant species differing in salt tolerance. Journal of Plant Physiology, 159: 137-146.
- Munns, R., 1993. *Physiological processes limiting plant growth in saline soil: some dogmas and hypotheses.* Plant, Cell and Environment, 16: 15-24.
- Munns, R., and James, R.A., 2003. Screening methods for salt tolerance: a case study with tetraploid wheat. Plant and Soil., 253: 201-218.

- Munns, R., James, A.J., Läuchli, A., 2006. Approaches to increasing the salt tolerance of wheat and other cereals. Journal of Experimental Botany, 57: 1025-1043.
- Munns, R., Schachtman, D.P., Condon, A.G., 1995. *The significance of a two-phase growth response to salinity in wheat and barley*. Australian Journal of Plant Physiology, 22: 561-569.
- Pakniyat, H.A., Kazemipour, A., and Mohammadi, A., 2003. Variation in salt tolerance of cultivated barley (Hordeum vulgare L.) and wild (H. spontaneum C. Koch) barley genotypes from Iran. Iran Agric. Res., 22: 45-62.
- Pitman, M.G., and Läuchli, A., 2002. Global impact of salinity and agricultural ecosystems. In: Läuchli

A., Lüttge U. (eds.), Salinity: environment – plants – molecules. Dordrecht: Kluwer: 3-20.

- SAS Institute,1985. *SAS users guide. Statistics*. Version 5 ed. SAS Inst., Cary, NC., USA.
- Schachtman, D.P, Munns, R, Whitecross, M.I., 1991. Variation of sodium exclusion and salt tolerance in Triticum tauschii. Crop Science, 31: 992-997.
- Taiz, L., and Zeiger, E., 2002. Plant physiology. Sunderland, Massachusetts Sinauer Associates, Inc., Publishers.
- Wilhelm, W.W., Bouzerzour, H., and Power, J.F., 1989. Soil disturbance-residue management effect on winter wheat growth and yield. Agron. J., 81: 581-588.